Protocol-at-a-Glance

QuickClean™ Enzyme Removal Resin Protocol-at-a-Glance (PT3159-2)

Please refer to the Certificate of Analysis for additional information.

Important notes on resin storage

- Do not autoclave or boil QuickClean.
- Protect QuickClean from long exposure to bright light.
- QuickClean is supplied as a 25% slurry.
- Do not allow QuickClean to dry out.

Important notes on resin use

- Stay within the optimal pH range for QuickClean (pH 5.0–8.0).
- Keep the concentration of salt in the extraction mixture under 0.3 N.
- After extraction with QuickClean, completely remove all traces of the blue resin (Step 7). Residual resin may carry traces of adsorbed proteins, and some enzymes retain their activity even when adsorbed to the resin.
- A centrifuge filtration device with a 0.45 µm pore size filter may be used to remove QuickClean resin from the sample. Follow the manufacturer's instructions for use of the filter.
- Do not re-use QuickClean.

PROTOCOL

1. Vigorously vortex the tube to resuspend QuickClean to homogeneity.
2. Add 1/10th volume of QuickClean to the mixture to be deproteinized in a 1.5 ml microcentrifuge tube. The final working concentration of Quick-Clean is 2.5%. Use a wide-bore tip to pipette the resin.
3. Vortex the mixture for 10–20 sec. If the DNA has a high molecular weight, do not vortex the sample. Instead, keep QuickClean in suspension by gently rotating or inverting the tube; alternatively, slowly pipet the solution up and down using a wide-bore pipette tip.
4. Centrifuge the tube at maximum speed (14,000 rpm) for 1 min to pellet protein/QuickClean complexes.
5. Transfer the supernatant to a fresh tube.
6. Repeat Steps 1–5 once or twice (see sample data on Certification of Analysis).
7. Completely remove remaining traces of QuickClean by centrifuging the supernatant one more time at 14,000 rpm for 1 min. Using a regular pipette tip, carefully transfer supernatant to a fresh tube.

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