# I. General Considerations

### A. Storage

Store at 4°C upon receipt. DO NOT FREEZE.

#### B. Stability

Stable for 1 year from date of receipt under proper storage conditions.

#### C. Required Materials

- Lenti-X Accelerator (Cat. Nos. 631256 or 631257)
- Magnetic Separator for Cell Culture (available separately as Cat. No. 631255, or combined with Lenti-X Accelerator in the Lenti-X Accelerator Starter Kit, Cat. No. 631254)
- Complete medium without antibiotics

## D. Cell Plating & Transduction Conditions

• Plate your cells one day prior to transduction. They should be 60–80% confluent on the day of transduction. Cultures that are less than 50% confluent at the time of transduction may lose viability.

**IMPORTANT: Antibiotic-free medium (no Penicillin-Streptomycin) is recommended during transduction to minimize any decrease in viability.** Please do not use any Penicillin G and Streptomycin (Pen-Strep) in the media during transduction. Pen-Strep may safely be added to the media after at least two passages, when the density of beads is typically lowered.

• The recommended amount of Lenti-X Accelerator (magnetic bead suspension) is typically sufficient to bind the number of viral particles produced using standard lentivirus production protocols. (1 x 10<sup>5</sup> to 1 x 10<sup>8</sup> IFU/ml). Table 1 provides the recommended range of bead and virus suspension volumes and cell densities for different sizes of culture vessel (plates and dishes).

**NOTE:** We recommend testing a few different bead volumes with a fixed virus dosage to achieve the highest transduction efficiency possible with your particular cell type.

Culture Vessel	Growth Area (cm <sup>2</sup> )	Volume of Medium (ml)	Cell Density	Volume of Beads (µl)	Volume of Virus (µl)
96-well plate	0.32	0.2	$0.04-0.1 \ge 10^5$	0.5-2	5-20
24-well plate	1.88	0.5	0.2–0.6 x 10 <sup>5</sup>	2-8	20-80
12-well plate	3.83	1	$0.4-1.2 \ge 10^5$	4–16	40-160
6-well plate	9.4	3	$1-3 \ge 10^5$	8-30	80-300
60-mm dish	21	5	$2-6 \ge 10^5$	16-64	160-640
100-mm dish	55	10	$5-15 \ge 10^5$	32-128	320-1280

Table I. Recommended Plating &	Transduction	<b>Conditions for Different</b>	Sizes of Culture	Vessel
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### II. Procedure

Perform all of the following steps under sterile conditions:

- 1. Mix Lenti-X Accelerator magnetic bead suspension by either pipetting up and down several times or by vortexing to achieve a uniform bead suspension.
- 2. Add the recommended volume (Table 1) to a sterile 1.5-ml microfuge tube.
- 3. Add the corresponding volume of virus-containing supernatant (Table 1) to the same tube and mix by gently pipetting up and down. Your virus may be resuspended in PBS or complete media.
- 4. Incubate the bead-virus mixture at room temperature or on ice for 20–30 min. During this incubation, tap the tube gently every 5 min to mix.

**NOTE:** The volume of magnetic bead suspension relative to that of virus suspension should be 10% or higher (see Table 1). A lower ratio requires a longer binding time.

- 5. Add the magnetic particles-virus mixture dropwise to the cells to be transduced. Rock the culture plate/dish gently to distribute the particles evenly across all the cells.
- 6. Place the culture plate/dish on top of the magnetic plate (i.e., Magnetic Separator for Cell Culture) for 5 min at room temperature or in a 37°C CO<sub>2</sub> incubator.
- 7. Change the media and then remove the culture from the magnetic plate. Cultivate cells under standard conditions. Any residual beads will be lost as the culture is passaged.

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