



	End of DD	BamHI		PvuII	MluI	NheI
659	AAA CCG GAA	GGA TCC	TCT AGT	CAG CTG	ACG CGT	GCT AGC
	TTT GGC CTT	CCT AGG	AGA TCA	GTC GAC	TGC GCA	CGA TCG
	NotI		HindIII			
	EagI	ClaI		EcoRV		
698	GCG GCC GCA	TCG ATA	AGC TTG	TCG ACG	ATA TCT	CCA
	CGC CGG CGT	AGC TAT	TCG AAC	AGC TGC	TAT AGA	GGT

pTRE-Cycle3 Vector Map and Multiple Cloning Site (MCS).

Description

pTRE-Cycle3 is a bidirectional, mammalian expression vector that lets you cycle the amount of your protein of interest in cells. Protein expression is tightly regulated by a bidirectional, tetracycline(Tet)-responsive promoter. Once expression is induced, protein levels can be rapidly reduced by simultaneously shutting down transcription and inducing rapid proteasomal degradation. This process can be reversed at any time, allowing the protein of interest to rapidly accumulate once again. In addition, the bidirectional promoter provides concurrent, Tet-regulated coexpression of the green fluorescent protein ZsGreen1.

pTRE-Cycle3 contains two main features that make such precise control over protein levels possible. First, expression of the gene of interest is tightly controlled by $P_{Tight-BI}$, a bidirectional, Tet-responsive promoter. $P_{Tight-BI}$ consists of two minimal CMV promoters ($P_{minCMV1}$ and $P_{minCMV2}$) and a modified Tet response element (TRE_{mod}) that consists of seven direct repeats of a 36 bp regulatory sequence containing the 19 bp tet operator sequence (*tetO*; 1). Second, the vector encodes a ProteoTuner™ destabilization domain (DD; 2). This domain is located between P_{Tight} and the multiple cloning site (MCS), allowing the addition of an N-terminal DD tag to your protein of interest. The DD tag causes the rapid degradation of any protein to which it is fused. This degradation can be prevented by the addition of Shield1 stabilizing ligand to the culture medium. Shield1 'shields' the fusion protein from proteasomal degradation, allowing the rapid accumulation of the tagged protein. When Shield1 is removed from the medium, the tagged protein is rapidly degraded.

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ZsGreen1, a human codon-optimized variant of the reef coral *Zoanthus sp.* green fluorescent protein (ZsGreen) that has been engineered for brighter fluorescence (3, 4), is positioned downstream of P_{minCMV2} . As a result, ZsGreen1 is coexpressed with the DD-tagged protein of interest. The vector also contains a pUC origin of replication and an ampicillin resistance gene (Amp^r) for propagation and selection in *E. coli*.

Use

pTRE-Cycle3 allows tightly regulated, doxycycline(Dox)-controlled coexpression of a DD-tagged protein of interest, and the fluorescent protein ZsGreen1. To create your DD-tagged protein of interest, your gene of interest must be cloned into the MCS in the same reading frame as the DD tag sequence. Dox-regulated expression of the proteins requires the presence of a tetracycline-controlled transcriptional activator, supplied by a stable Tet-On[®] Advanced or Tet-Off[®] Advanced cell line that can be created with our Tet-On Advanced or Tet-Off Advanced Inducible Gene Expression Systems (Cat. Nos. 630930 and 630934). These systems provide the inducible gene expression strategy of Gossen & Bujard, with major improvements described by Urlinger, *et al.* (5–9).

The effects of Dox and Shield1 are concentration-dependent and reversible. Therefore, it is possible to fine-tune: a) the amount of both the DD-tagged protein of interest and ZsGreen1 present in the cells by adjusting the concentration of Dox in the medium; or b) the amount of just the DD-tagged protein of interest by adjusting the concentration of Shield1 in the medium.

Dox-regulated expression of ZsGreen1 allows the use of fluorescence microscopy or flow cytometry to easily monitor and/or select cells expressing the gene of interest (ZsGreen1 has an excitation maximum of 493 nm and an emission maximum of 505 nm).

Location of Features

- $P_{\text{Tight-BI}}$ (bidirectional, Tet-responsive promoter):
 TRE_{mod} (modified Tet-response element): 3–252
 P_{minCMV1} (minimal CMV promoter 1): 258–317
 P_{minCMV2} (minimal CMV promoter 2): 3796–3864 (complementary)
- DD (ProteoTuner destabilization domain): 344–667
- MCS (multiple cloning site): 668–729
- SV40 polyA signal: 741–928
- ColE1 origin of replication: 1104–1703
- Amp^r (ampicillin resistance gene; β -lactamase): 1865–2860 (complementary)
- SV40 polyA signal: 2861–3048 (complementary)
- ZsGreen1 (*Zoanthus sp.* green fluorescent protein): 3083–3778 (complementary)

Propagation in *E. coli*

- Recommended host strain: DH5 α [™], HB101, and other general purpose strains.
- Selectable marker: plasmid confers resistance to ampicillin (100 μ g/ml) in *E. coli* hosts.
- *E. coli* replication origin: ColE1
- Plasmid incompatibility group: pMB1/ColE1

Excitation and Emission Maxima of ZsGreen1

- Excitation maximum = 493 nm
- Emission maximum = 505 nm

References

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5. Gossen, M. & Bujard, H. (1992) *Proc. Natl. Acad. Sci USA* **89**(12):5547–5551.
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8. Inducible Gene Expression Systems (January 2007) *Clontechniques XXII*(1):1–2.
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Note: The vector sequence was compiled from information in the sequence databases, published literature, and other sources, together with partial sequences obtained by Clontech. This vector has not been completely sequenced.

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