



Restriction Map and Multiple Cloning Site (MCS) of pZsGreen1-C1. All sites shown are unique. The *Xba* I site (*) is methylated in the DNA provided by CLONTECH. If you wish to digest the vector with these enzymes, you will need to transform the vector into a *dam*⁻ host and make fresh DNA.

Description

pZsGreen1-C1 encodes a human codon-optimized variant of wild-type *Zoanthus* sp. green fluorescent protein, ZsGreen1 (1). The ZsGreen1 coding sequence contains a series of silent base-pair changes, which correspond to human codon-usage preferences, for optimal expression in mammalian cells (2). Additionally, an upstream sequence—located just 5' to the ZsGreen1 start codon—has been converted to a Kozak consensus translation initiation site (3) to further increase the translation efficiency in eukaryotic cells. A single amino acid substitution (Asn-65 to Met) has been made to enhance the emission characteristics of ZsGreen1 (excitation maximum = 496 nm; emission maximum = 506 nm).

The multiple cloning site (MCS) in pZsGreen1-C1 is positioned between the ZsGreen1 coding sequence and a pair of SV40 polyadenylation signals (SV40 poly A). Thus, genes cloned into the MCS will be expressed as fusions to the C-terminus of ZsGreen1 if they are in the same reading frame as ZsGreen1 and there are no intervening stop codons. Expression of ZsGreen1 is driven by the cytomegalovirus immediate-early promoter ($P_{CMV IE}$). The SV40 poly A signals downstream of the MCS direct proper processing of the 3' end of ZsGreen1 mRNA.

The vector backbone contains an SV40 origin (SV40 ori) for replication in mammalian cells that express the SV40 T-antigen, a pUC origin of replication for propagation in *E. coli*, and an f1 origin for single-stranded DNA production. A neomycin resistance cassette—consisting of the SV40 early promoter (P_{SV40_e}), the neomycin/kanamycin resistance gene of Tn5 (Neo^r/Kan^r), and polyadenylation signals from the Herpes simplex virus thymidine kinase (HSV TK poly A) gene—allows stably transfected eukaryotic cells to be selected using G418 (4). A bacterial promoter (P) upstream of this cassette drives expression of the Neo^r/Kan^r gene in *E. coli* hosts, which can be selected with kanamycin.



United States/Canada
800.662.2566

Asia Pacific
+1.650.919.7300

Europe
+33.(0)1.3904.6880

Japan
+81.(0)77.543.6116

Clontech Laboratories, Inc.
A Takara Bio Company
1290 Terra Bella Ave.
Mountain View, CA 94043
Technical Support (US)
E-mail: tech@clontech.com
www.clontech.com

(PR641614; published 24 April 2006)

Use

Fusions to the C terminus of ZsGreen1 retain the fluorescent properties of the native protein allowing the localization of the fusion protein *in vivo*. The target gene should be cloned into pZsGreen1-C1 so that it is in frame with the ZsGreen1 coding sequence, with no intervening, in-frame stop codons. The recombinant pZsGreen1-C1 vector can be transfected into mammalian cells using any standard transfection method. If required, stable transformants can be selected using G418 (available from Clontech; Cat. Nos. 631307 & 631308). We recommend selecting mammalian cell cultures in 500–1,300 µg/ml G418, depending on the cell line. Be sure to establish a kill curve for each cell line and each lot of G418 to determine optimal selection concentration. Unmodified (i.e., non-recombinant) pZsGreen1-C1 can also be used simply to express ZsGreen1 in a cell line of interest (e.g., as a transfection marker).

Location of features

- Human cytomegalovirus (CMV) immediate early promoter: 1–589
Enhancer region: 59–465; TATA box: 554–560
Transcription start point: 583
C→G mutation to remove *Sac* I site: 569
- *Zoanthus* sp. green fluorescent protein (ZsGreen1) coding sequence
Kozak consensus translation initiation site: 606–616
Start codon (ATG): 613–615
Asn-65 to Met mutation (A→T, C→G): 809, 810
- Multiple Cloning Site (MCS): 1315–1383
Stop codon: 1384–1386
- SV40 early mRNA polyadenylation signal
Polyadenylation signals: 1526–1531 & 1555–1560; mRNA 3' ends: 1564 & 1576
- f1 single-strand DNA origin: 1623–2078 (Packages the noncoding strand of ZsGreen1.)
- Bacterial promoter for expression of Kan^r gene
–35 region: 2140–2145; –10 region: 2163–2168
Transcription start point: 2175
- SV40 origin of replication: 2419–2554
- SV40 early promoter
Enhancer (72-bp tandem repeats): 2252–2323 & 2324–2395
21-bp repeats: 2399–2419, 2420–2440 & 2442–2462
Early promoter element: 2475–2481
Major transcription start points: 2471, 2509, 2515 & 2520
- Kanamycin/neomycin resistance gene
Neomycin phosphotransferase coding sequences:
Start codon (ATG): 2603–2605; stop codon: 3395–3397
G→A mutation to remove *Pst* I site: 2785
C→A (Arg to Ser) mutation to remove *Bss*H II site: 3131
- Herpes simplex virus (HSV) thymidine kinase (TK) polyadenylation signal
Polyadenylation signals: 3633–3638 & 3646–3651
- pUC plasmid replication origin: 3982–4625

Propagation in *E. coli*

- Suitable host strains: DH5 α , HB101, and other general purpose strains. Single-stranded DNA production requires a host containing an F plasmid such as JM109 or XL1-Blue.
- Selectable marker: plasmid confers resistance to kanamycin (50 µg/ml) to *E. coli* hosts.
- *E. coli* replication origin: pUC
- Copy number: ~500
- Plasmid incompatibility group: pMB1/Col E1

References

1. Matz, M. V., *et al.* (1999) *Nature Biotech.* **17**:969–973.
2. Haas, J., *et al.* (1996) *Curr. Biol.* **6**:315–324.
3. Kozak, M. (1987) *Nucleic Acids Res.* **15**:8125–8148.
4. Gorman, C. (1985). In *DNA Cloning: A Practical Approach, Vol. II*. Ed. D.M. Glover. (IRL Press, Oxford, U.K.) pp. 143–190.

Note: The attached sequence file has been compiled from information in the sequence databases, published literature, and other sources, together with partial sequences obtained by Clontech. This vector has not been completely sequenced.

Notice to Purchaser

This product is intended to be used for research purposes only. It is not to be used for drug or diagnostic purposes nor is it intended for human use. Clontech products may not be resold, modified for resale, or used to manufacture commercial products without written approval of Clontech Laboratories, Inc.

This product is the subject of pending U.S. patents.

Not-For-Profit Entities: Orders may be placed in the normal manner by contacting your local representative or Clontech Customer Service at 650.919.7300. At its discretion, Clontech grants Not-For-Profit Entities a non-exclusive, personal, limited license to use this product for non-commercial life science research use only. Such license specifically excludes the right to sell or otherwise transfer this product, its components or derivatives thereof to third parties. No modifications to the protein coding sequence may be made without express written permission from Clontech. Any other use of this product requires a license from Clontech. For license information, please contact a licensing representative by phone at 650.919.7320 or by e-mail at licensing@clontech.com.

For-Profit Entities wishing to use this product are required to obtain a license from Clontech. For license information, please contact a licensing representative by phone at 650.919.7320 or by e-mail at licensing@clontech.com.

Clontech, Clontech logo and all other trademarks are the property of Clontech Laboratories, Inc. Clontech is a Takara Bio Company. ©2005