

**qPCR Human Reference Total RNA** is a mixture of total RNAs from multiple adult human tissues chosen to represent a broad range of expressed genes. It is a reliable reference standard for the accurate and reproducible comparison of gene expression data using real-time quantitative PCR (qPCR). Our qPCR Reference RNA may also be used as a source of positive control templates for validating qPCR primer designs.

This Protocol-at-a-Glance provides a sample qPCR protocol using our TB Green® Advantage® qPCR Premix (Cat. No. 639676) to be used once the individual researcher has generated first-strand reference cDNA with the protocol of their choice.

## I. List of Components

- qPCR Human Reference Total RNA (Store at  $-70^{\circ}\text{C}$ )

## II. Additional Materials Required

For qPCR amplification:

- TB Green Advantage qPCR Premix (Cat. No. 639676)
- qPCR primers
- Supplies and equipment for performing qPCR
- First strand cDNA from an RT reaction, with a minimal concentration of 50 ng/ $\mu\text{l}$  input RNA.

## III. Protocol: qPCR Amplification

This protocol is a guide to performing qPCR using the first-strand reference cDNA and TB Green Advantage qPCR Premix.

1. If needed, dilute the reaction mixture from the reverse transcriptase reaction down to a final concentration of 10 ng/ $\mu\text{l}$  input RNA.
2. Prepare a series of six cDNA reference standards, following the guidelines presented in Table I. Include a no-template control (40  $\mu\text{l}$  RNase-free  $\text{H}_2\text{O}$ ) as an additional standard (Standard No. 7).

**Table 1. Guidelines for serial dilution**

Standard No.	RNA concentration	Aliquot	Volume of RNase-free $\text{H}_2\text{O}$ to add
1	10 ng/ $\mu\text{l}$	None	None
2	2 ng/ $\mu\text{l}$	10 $\mu\text{l}$ Standard No. 1	40 $\mu\text{l}$
3	0.4 ng/ $\mu\text{l}$	10 $\mu\text{l}$ Standard No. 2	40 $\mu\text{l}$
4	0.08 ng/ $\mu\text{l}$	10 $\mu\text{l}$ Standard No. 3	40 $\mu\text{l}$
5	0.016 ng/ $\mu\text{l}$	10 $\mu\text{l}$ Standard No. 4	40 $\mu\text{l}$
6	0.0032 ng/ $\mu\text{l}$	10 $\mu\text{l}$ Standard No. 5	40 $\mu\text{l}$
7	0 ng/ $\mu\text{l}$	N/A	40 $\mu\text{l}$

3. Prepare a qPCR Master Mix for all reaction wells, plus one additional well. Combine the following components in the order shown:

<u>per rxn</u>	
X $\mu\text{l}$	RNase-free $\text{H}_2\text{O}$
12.5 $\mu\text{l}$	TB Green Advantage qPCR Premix (2X)
Y $\mu\text{l}$	Primer mix (20–400 nM)*
20 $\mu\text{l}$	Total volume

\* The amount varies according to application.

- Mix well by vortexing and spin the tube briefly in a microcentrifuge.
- Aliquot 20 µl of qPCR Master Mix into each well of a 96-well optical PCR plate.
- Add 5 µl of Standard No. 1 (from Step 1, Table I) to a well. Repeat, adding the next Standard in the series to each subsequent well, to obtain a qPCR reaction for each Standard. Seal the plate using optical strip caps.
- Begin thermal cycling using the parameters optimized for your primers.

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This document has been reviewed and approved by the Quality Department.