TaKaRa LA Taq® DNA Polymerase

Code No. RR002M Size: 250 units

Shipping at -20° C Store at -20° C

Supplied Reagents:

10X LA PCR Buffer II (25 mM Mg²⁺ plus) 1 ml dNTP Mixture (2.5 mM each) 800 μ I

Lot No.

Conc.: units/ μ I Volume: μ I

Expiration Date:

Storage Buffer: 20 mM Tris-HCl (pH8.0)

100 mM KCl 0.1 mM EDTA 1 mM DTT 0.5% Tween 20 0.5% Nonidet P-40 50% Glycerol

Supplied dNTP Mixture (2.5 mM each)

The dNTP Mixture is ready for use in PCR without dilution.

Form : Dissolved in water (sodium salts), pH7 - 9

Purity : ≧ 98% for each dNTP

Unit definition:

One unit is the amount of enzyme that incorporates 10 nmol of dNTPs into acid-insoluble products in 30 minutes at 74° C with activated salmon sperm DNA as the template-primer.

Reaction mixture for unit definition:

25 mM TAPS (pH9.3 at 25°C)

50 mM KCl 2 mM MgCl₂ 0.1 mM DTT

200 μM each dATP·dGTP·dCTP

100 μ M [3 H]-dTTP

0.25 mg/ml activated salmon sperm DNA

Purity: Nicking, endonuclease, and exonuclease activity were not detected after incubation of 0.6 μ g of supercoiled pBR322 DNA, 0.6 μ g of λ DNA, or 0.6 μ g of λ -Hind III digest with 10 units of this enzyme for 1 hour at 74°C.

Applications: For DNA amplification by PCR. This enzyme is optimized for long range PCR (>15 kb fragments).

PCR products:

As most PCR products amplified with $TaKaRa\ LA\ Taq$ DNA polymerase have one A at the 3'-termini, the obtained PCR products can be directly used for cloning into T-Vectors. When cloning long products (>5 kb) into T-Vectors, the cloning efficiency may be low. It is also possible to clone the product in blunt-end vectors after blunting and phosphorylation of the end.

PCR test:

Good performance in PCR was confirmed by amplification of a 35 kb fragment from λ DNA template.

Good performance in PCR was confirmed by amplification of the β -globin gene (17.5 kb) using human genomic DNA template.

General reaction mixture for PCR (50 μ l reaction volume):

<i>TaKaRa LA Taq</i> (5 units/ μ l)	0.5 μΙ
10X LA PCR Buffer II (Mg ²⁺ plus)	5 μΙ
dNTP Mixture (2.5 mM each)	8 μΙ
Template	<1 μg
Primer 1	$0.2 - 1.0 \mu M$ (final conc.)
Primer 2	$0.2 - 1.0 \mu M$ (final conc.)
Sterile distilled water	up to 50 μl

PCR conditions (example): Amplification of a 17.5 kb DNA fragment

94°C 1 min. 98°C 10 sec. 68°C 15 min. 72°C 10 min. 30 cycles

(Note) Denaturation conditions vary depending on the thermal cycler and tubes used for PCR. Denaturation for 5 - 10 sec. at 98°C or 20 - 30 sec. at 94°C is recommended.

< Cool Start Method >

The "Cool Start Method" provides more accurate amplification and minimizes amplification of nonspecific products. This simple method does not require specialized enzymes or additional reagents.

Cool Start Method Protocol

- 1) Keep all reagents on ice until use.
- 2) Prepare the reaction mixture on ice. *1,2
 - *1: Order of reagent addition does not influence results.
 - *2: Results will not be affected by leaving the mixture on ice for up to 30 min. before thermal cycling.
- 3) Set the thermal cycler with the designated program. *3
- * 3: PCR conditions do not need to be changed for the Cool Start Method. 4) Set the tubes in the thermal cycler and start cycling immediately.

NOTICE TO PURCHASER: LIMITED LICENSE

[P1] PCR Notice

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[M57] LA Technology

This product is covered by the claims 6-16 outside the U.S. corresponding to the expired U.S. Patent No. 5,436,149.

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Note

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