

## His60 Ni Superflow Cartridges

**Catalog No.**  
635675

**Amount**  
5 x 1 ml cartridges

**Lot Number**  
Specified on product label.

### Description

Prepacked cartridges, each containing 1 ml of His60 Ni Superflow Resin, for the purification of recombinant his-tagged proteins. The cartridges are suitable for automated purification using medium-pressure systems such as ÄKTA FPLC or manual, syringe-based purification. Proteins can be purified under native or denaturing conditions. Each column can be reused up to 5 times to purify different preparations of the same protein.

### Package Contents

- 5 His60 Ni Superflow Cartridges (1 ml each)
- 5 Top Caps
- 5 Snap-off End Caps

### Storage Conditions

- Store all components at 4°C.

### Shelf Life

- 1 year from date of receipt under proper storage conditions.

### Storage Buffer

- Nonbuffered 20% ethanol

### Shipping Conditions

- Room temperature

### Product Documents

Documents for Clontech® products are available for download at [www.clontech.com/manuals](http://www.clontech.com/manuals).

The following document applies to this product:

His60 Ni Superflow Cartridges User Manual (PT5143-1)

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#### Clontech Laboratories, Inc.

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(030816)

## Quality Control Data

- **Metal loading analysis:**

Each lot of His60 Ni Superflow Cartridges is guaranteed to have a nickel metal loading of  $\geq 25$   $\mu\text{mol/ml}$  of bed volume. The amount of bound nickel is determined by atomic spectroscopy.

- **Functional test:**

The His60 Ni Superflow Resin in the His60 Ni Superflow Cartridges was functionally tested using his-tagged GFPuv and the His60 Ni Buffer Set (Cat. No. 635665). One ml of His60 Ni Superflow Resin was saturated with a bacterial his-tagged GFPuv lysate in xTractor™ Buffer. The resin was washed two times with 5 ml of His60 Ni Equilibration Buffer (pH 7.4) and then two times with 5 ml of Wash Buffer (pH 7.4). Bound protein was eluted three times with 1 ml of His60 Ni Elution Buffer (pH 7.4). All eluted fractions were analyzed by electrophoresis on a 12% SDS polyacrylamide gel to verify the purity of the eluted protein. A strong his-tagged GFPuv protein band was evident in at least one lane containing eluted sample.

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