

## pDD-ZsGreen1 Reporter

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Catalog No.	Amount	Lot Number
632195 (Not sold separately)	20 µg	Specified on product label.

### Product Information

pDD-ZsGreen1 Reporter is sold as part of the DD-ZsGreen1 Reporter System (Cat. No. 632192). pDD-ZsGreen1 Reporter is a promoterless vector that can be used to monitor transcription from different promoters and promoter/enhancer combinations inserted into the multiple cloning site (MCS). The gene downstream of the MCS encodes the bright green fluorescent protein ZsGreen1, tagged at its N-terminus with the ProteoTuner™ destabilization domain (DD; 1). In the absence of the Shield1 ligand, the DD tag induces rapid degradation of the fluorescent reporter, minimizing any background caused by leaky promoters; but upon addition of Shield1 at the time of promoter activation, the DD-tagged reporter molecules are stabilized, increasing the signal-to-noise ratio.

### Package Contents

- 20 µg pDD-ZsGreen1 Reporter

### Storage Conditions

- Store at -20°C
- Spin briefly to recover contents
- Avoid repeated freeze/thaw cycles

### Shelf Life

- 1 year from date of receipt under proper storage conditions.

### Storage Buffer

- 10 mM Tris-HCl (pH 8.0)
- 1 mM EDTA (pH 8.0)

### Concentration

- 500 ng/µl

### Shipping Conditions

- Dry ice (-70°C)

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#### Clontech Laboratories, Inc.

A Takara Bio Company  
1290 Terra Bella Avenue, Mountain View, CA 94043, USA  
U.S. Technical Support: [tech@clontech.com](mailto:tech@clontech.com)

United States/Canada	Asia Pacific	Europe	Japan
800.662.2566	+1.650.919.7300	+33.(0)1.3904.6880	+81.(0)77.543.6116
(PA124361)			

## Product User Manuals

User manuals for Clontech products are available for download at [www.clontech.com/manuals](http://www.clontech.com/manuals).

The following user manuals apply to this product:

- DD-Fluorescent Protein Reporter Systems Protocol-At-A-Glance (PT4088-2)
- ProteoTuner Systems User Manual (PT4039-1)

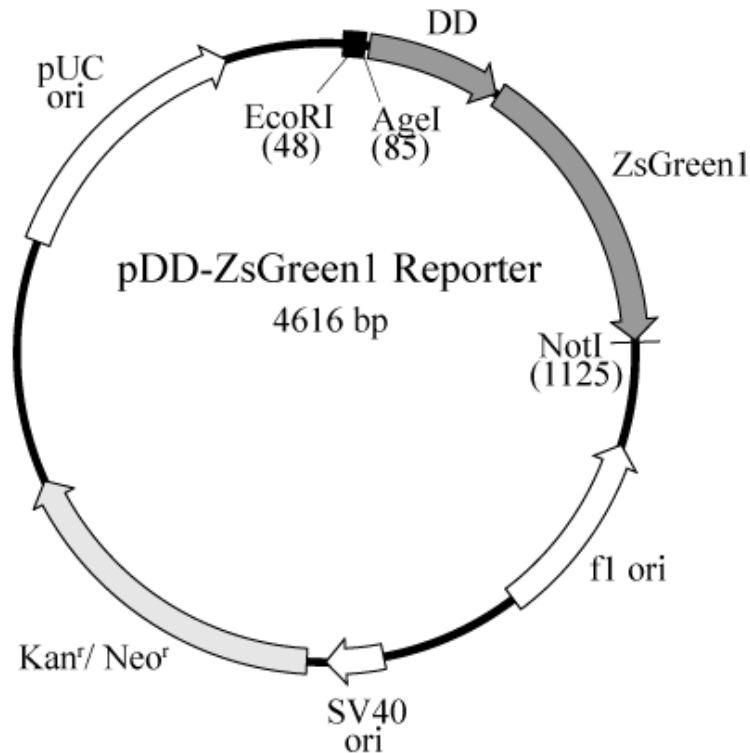


Figure 1. pDD-ZsGreen1 Reporter vector map.

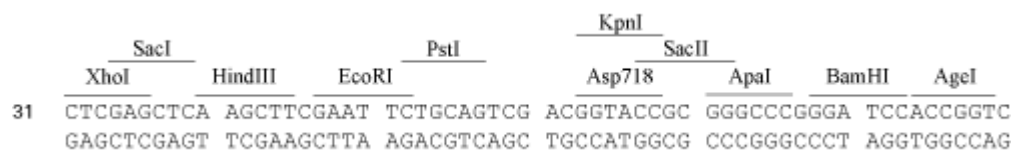


Figure 2. pDD-ZsGreen1 Reporter vector multiple cloning site.

## Description

pDD-ZsGreen1 Reporter is a promoterless reporter vector that allows the functional analysis of different promoters and promoter/enhancer combinations inserted into its multiple cloning site (MCS). The vector encodes the reporter protein DD-ZsGreen1, a ligand-dependent, destabilized bright green fluorescent protein that minimizes background fluorescence from leaky promoters. This reporter can be used to monitor promoter activity in live cells and *in vivo*. A promoter must be cloned into the MCS, located upstream of the DD-ZsGreen1 coding sequence. Without the addition of a functional promoter, the vector will not express DD-ZsGreen1.

## DD-ZsGreen1 Reporter

ZsGreen1 (excitation and emission maxima: 493 and 505 nm, respectively) is a human codon-optimized variant of the reef coral *Zoanthus* sp. green fluorescent protein (ZsGreen) that has been engineered for brighter fluorescence (2, 3). DD-

ZsGreen1 is a modified version of ZsGreen1 that is tagged on its N-terminus with the ProteoTuner DD, which causes rapid, proteasomal degradation of DD-ZsGreen1 (1). However, when the membrane-permeant, stabilizing ligand Shield1 is added to the medium, it binds to the DD and prevents degradation of the DD-ZsGreen1 reporter protein, thereby causing it to accumulate inside the cell.

In the absence of Shield1, the DD causes the degradation of any DD-ZsGreen1 reporter protein produced prior to promoter activation, thus minimizing background fluorescence caused by leaky promoters. To analyze promoter activity, the inducer of choice is added to the medium along with Shield1, which effectively stabilizes the reporter protein, allowing it to accumulate. As a result, only the reporter molecules expressed during promoter induction will contribute to the fluorescence signal, providing a considerably higher signal-to-noise ratio than that obtained with non-destabilized or constitutively destabilized reporter systems.

The promoter's activity level can be directly correlated to the fluorescence level.

## Vector Elements

The vector backbone contains an SV40 origin for replication in mammalian cells expressing the SV40 large T antigen, a pUC origin of replication for propagation in *E. coli*, and an f1 origin for single-stranded DNA production. pDD-ZsGreen1 can be transfected into mammalian cells using any standard transfection method.

## Antibiotic Selection

A neomycin-resistance cassette (Neo<sup>r</sup>) allows stably transfected eukaryotic cells to be selected using G418 (4). This cassette consists of the SV40 early promoter, a Tn5 kanamycin/neomycin resistance gene, and herpes simplex virus thymidine kinase (HSV TK) polyadenylation signals. A bacterial promoter upstream of the cassette expresses kanamycin resistance in *E. coli*.

## Location of Features

- MCS (multiple cloning site): 31–89
- DD-ZsGreen1  
Start codon (ATG): 97–99; Stop codon: 1114–1116  
DD (FKBP-L106P destabilization domain; 1): 97–420  
ZsGreen1 (*Zoanthus* sp. green fluorescent protein): 427–1119
- f1 origin of replication (for packaging the noncoding strand of DD-ZsGreen1): 1372–1827 (complementary)
- SV40 origin of replication: 2168–2306
- Kan<sup>r</sup>/Neo<sup>r</sup> (kanamycin/neomycin resistance gene)  
Neomycin phosphotransferase coding sequences: 2352–3146
- pUC origin of replication: 3731–4374

## Additional Information

### Propagation in *E. coli*

- Recommended host strains: DH5 $\alpha$ , HB101, and other general purpose strains. Single-stranded DNA production requires a host containing an F plasmid such as JM109 or XL1-Blue.
- Selectable marker: plasmid confers resistance to kanamycin (50  $\mu$ g/ml) in *E. coli* hosts.

# Certificate of Analysis

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- *E. coli* replication origin: pUC
- Copy number: high
- Plasmid incompatibility group: pMB1/ColE1

## Excitation and Emission Maxima of ZsGreen1

- Excitation: 493 nm
- Emission: 505 nm

## References

1. Banaszynski, L. *et al.* (2006) *Cell* **126**(5):995–1004.
2. Matz, M. V. *et al.* (1999) *Nature Biotech.* **17**(10):969–973.
3. Haas, J. *et al.* (1996) *Curr. Biol.* **6**(3):315-324
4. Gorman, C. (1985) In *DNA Cloning: A Practical Approach, Vol. II*. Ed. D. M. Glover. (IRL Press, Oxford, U.K.), pp. 143–190.

## Quality Control Data

### Plasmid Identity & Purity

- Digestion with the indicated restriction enzymes produced fragments of the indicated sizes on a 0.8% agarose/EtBr gel:

Enzymes	Fragments (kb)
EcoRI	4.6
AgeI & NotI	1.0 & 3.6

- Vector identity was confirmed by sequencing.
- $A_{260}/A_{280}$ : 1.8–2.0

*Note: The vector sequence was compiled from information in the sequence databases, published literature, and other sources, together with partial sequences obtained by Clontech. This vector has not been completely sequenced.*

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### LICENSING STATEMENTS:

The RCFP's (including DsRedExpress and DsRedExpress2) are covered by one or more of the following U.S. Patent Nos. 7,166,444; 7,157,565; 7,217,789; 7,338,784; 7,338,783; 7,537,915 6,969,597, 7,150,979 and 7,442,522.

Patent Pending.

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This document has been reviewed and approved by the Clontech Quality Assurance Department.

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### Clontech Laboratories, Inc.

A Takara Bio Company  
1290 Terra Bella Avenue, Mountain View, CA 94043, USA  
U.S. Technical Support: [tech@clontech.com](mailto:tech@clontech.com)

**United States/Canada**  
800.662.2566

**Asia Pacific**  
+1.650.919.7300

**Europe**  
+33.(0)1.3904.6880

**Japan**  
+81.(0)77.543.6116