

## pTRE-CellCycle Vector

**Catalog No.**

631466

**Amount**

10 µg

**Lot Number**

Specified on product label.

### Description

pTRE-CellCycle is a bidirectional, tetracycline (Tet)-inducible expression vector that lets you inducibly express fluorescent, ubiquitination-based, cell-cycle indicators (FUCCI; Sakaue-Sawano et al. 2008) in order to monitor all phases of the cell cycle. The vector allows you to identify cells that are either in G1 phase or transitioning between S, G2, and M phases. pTRE-CellCycle is intended for use with any Tet-On® or Tet-Off® Expression System.

### Package Contents

- 20 µl pTRE-CellCycle Vector (500 ng/µl)

### Storage Conditions

- Store plasmids at –20°C.
- Spin briefly to recover contents.
- Avoid repeated freeze/thaw cycles.

### Expiration Date

- Specified on product label.

### Storage Buffer

- 10 mM Tris-HCl (pH 8.0), 1 mM EDTA (pH 8.0)

### Shipping Conditions

- Dry ice

### Product Documents

Documents for our products are available for download at [takarabio.com/manuals](http://takarabio.com/manuals)

The following documents apply to this product:

- Tet-On Advanced Inducible Gene Expression System User Manual
- Tet-Off Advanced Inducible Gene Expression System User Manual

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## Vector Information

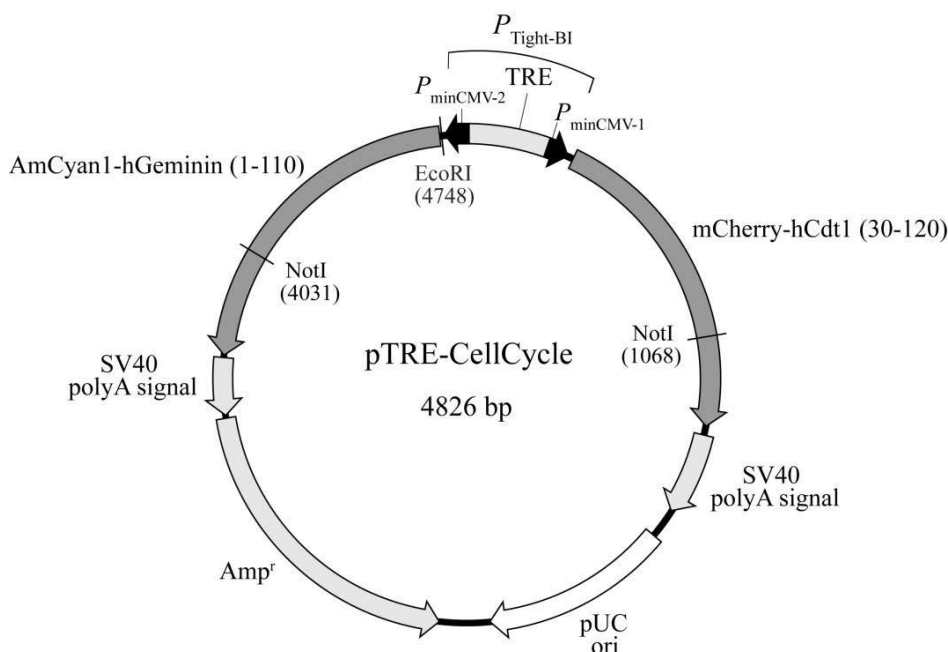


Figure 1. pTRE-CellCycle Vector Map.

## Description

pTRE-CellCycle is a bidirectional Tet-inducible expression vector that can be used to inducibly express the cell cycle indicators hGeminin (1–110), tagged with the cyan fluorescent protein AmCyan1, and hCdt1 (30–120), tagged with the red fluorescent protein mCherry (McGarry and Kirschner 1998; Nishitani, Lygerou, and Nishimoto 2004). Both fusions can be induced simultaneously in our Tet-On and Tet-Off cell lines.

The AmCyan1-hGeminin (1–110) fusion is present at high levels during the S, G2 and M phases of the cell cycle, whereas the mCherry-hCdt1 (30–120) fusion is present at high levels during the G1 phase of the cell cycle. AmCyan1 is a human-codon optimized variant of the wild type *Anemonia majano* cyan fluorescent protein (excitation and emission maxima of 458 nm and 489 nm, respectively; Shaner et al. 2004), and mCherry is a mutant fluorescent protein derived from the tetrameric *Discosoma* sp. red fluorescent protein DsRed (excitation and emission maxima of 587 nm and 610 nm, respectively). Because both fusions are expressed together, the presence and absence of cyan and red fluorescence can be used to monitor all phases of the cell cycle.

Expression of both fusion proteins is tightly controlled by  $P_{Tight-BI}$ , a bidirectional, Tet-responsive promoter.  $P_{Tight-BI}$  consists of a tetracycline response element (TRE) flanked by two minimal CMV promoters ( $P_{minCMV-1}$  and  $P_{minCMV-2}$ ). The TRE is recognized and bound by a Tet-On or Tet-Off transactivator in the presence or absence of doxycycline (Dox), respectively (Gossen and Bujard 1992; Gossen et al. 1995). Thus, expression of both genes is responsive to the transactivators in the Tet-On and Tet-Off systems. pTRE-CellCycle should be cotransfected with the Linear Hygromycin Marker (Takara Bio, Cat. No. 631625, not included) or Linear Puromycin Marker (Takara Bio, Cat. No. 631626, not included).

## Location of Features

- P<sub>Tight-BI</sub> (bidirectional, Tet-responsive promoter):  
TRE (Tet response element): 3–252  
P<sub>minCMV-1</sub> (minimal CMV promoter 1): 258–317  
P<sub>minCMV-2</sub> (minimal CMV promoter 2): 4758–4826 (complementary)
- *mCherry::hCdt1* (30–120) (mCherry and hCdt1 gene fusion): 341–1351
- SV40 polyA signal: 1374–1574
- pUC19 origin of replication: 1737–2336
- *Amp<sup>r</sup>* (ampicillin resistance gene;  $\beta$ -lactamase): 2497–3493 (complementary)
- SV40 polyA signal: 3480–3683 (complementary)
- *AmCyan1::hGeminin* (1–110) (AmCyan1 and hGeminin gene fusion): 3694–4740 (complementary)

## Additional Information

Dox-regulated expression of the proteins requires the presence of a tetracycline-controlled transcriptional activator, supplied by a stable Tet-On Advanced or Tet-Off Advanced cell line that can be created with our Tet-On Advanced or Tet-Off Advanced Inducible Gene Expression Systems (Takara Bio, Cat. Nos. 630930 and 630934). These systems provide the inducible gene expression strategy of Gossen & Bujard 1992, with major improvements described by Urlinger et al. 2000.

**NOTE:** Overexpression of the fusion proteins could lead to insufficient proteosomal degradation, which could prevent effective monitoring of the cell cycle. Therefore, we recommend either (i) optimizing gene transfer conditions to avoid overexpression of the fusion proteins, or (ii) selecting cells that express acceptable amounts of the fusion proteins.

## Propagation in *E. coli*

- Recommended host strain: Stellar<sup>TM</sup> Competent Cells
- Selectable marker: plasmid confers resistance to ampicillin (100  $\mu$ g/ml) in *E. coli* hosts.
- *E. coli* replication origin: pUC

## Excitation and emission maxima of AmCyan1

- Excitation maximum = 458 nm
- Emission maximum = 489 nm

## Excitation and emission maxima of mCherry

- Excitation maximum = 587 nm
- Emission maximum = 610 nm

## References

- Gossen, M. *et al.* Transcriptional activation by tetracyclines in mammalian cells. *Science* **268**, 1766–9 (1995).
- Gossen, M. & Bujard, H. Tight control of gene expression in mammalian cells by tetracycline-responsive promoters. *Proc. Natl. Acad. Sci. U. S. A.* **89**, 5547–51 (1992).
- McGarry, T. J. & Kirschner, M. W. Geminin, an inhibitor of DNA replication, is degraded during mitosis. *Cell* **93**, 1043–53 (1998).

# Certificate of Analysis

Cat. No. 631466

## pTRE-CellCycle Vector

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Nishitani, H., Lygerou, Z. & Nishimoto, T. Proteolysis of DNA replication licensing factor Cdt1 in S-phase is performed independently of geminin through its N-terminal region. *J. Biol. Chem.* **279**, 30807–16 (2004).

Sakaue-Sawano, A. *et al.* Visualizing Spatiotemporal Dynamics of Multicellular Cell-Cycle Progression. *Cell* **132**, 487–498 (2008).

Shaner, N. C. *et al.* Improved monomeric red, orange and yellow fluorescent proteins derived from *Discosoma* sp. red fluorescent protein. *Nat. Biotechnol.* **22**, 1567–72 (2004).

Urlinger, S. *et al.* Exploring the sequence space for tetracycline-dependent transcriptional activators: Novel mutations yield expanded range and sensitivity. *Proc. Natl. Acad. Sci. U. S. A.* **97**, 7963–7968 (2000).

## Quality Control Data

### Plasmid Identity & Purity

- Digestion with the indicated restriction enzymes produced fragments of the indicated sizes on a 0.8% agarose/EtBr gel:

Vector	Enzymes	Fragment Sizes
pTRE-CellCycle Vector	EcoRI	4.8 kb
	NotI	1.9 & 3.0 kb

- Vector identity was confirmed by sequencing.
- A<sub>260</sub>/A<sub>280</sub>: 1.8–2.0

It is certified that this product meets the above specifications, as reviewed and approved by the Quality Department.

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# Notice to Purchaser



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