

## pET6xHN Expression Vector Set (In-Fusion® Ready)

<b>Catalog No.</b>	<b>Amount</b>	<b>Lot Number</b>
631433 (Not sold separately) Sold as a part of 631428 & 631429	Each	Specified on product label.

### Description

The pET6xHN Expression Vector Set (In-Fusion Ready) allows you to use In-Fusion cloning technology to express your protein of interest with an N- or C-terminal 6xHN tag in *E. coli*. The vector set contains IPTG-inducible, pET-based vectors that have been prelinearized to allow In-Fusion cloning with the included In-Fusion HD Enzyme Premix. Each vector contains a T7/*lac* promoter for high-level expression of his-tagged proteins, which can be easily prepped for exceptional purity with our TALON® cobalt (Co) resins, or for standard purity and high yield with our high-capacity His60 nickel resins. The vector set also includes a control vector that expresses an N-terminal, 6xHN-tagged GFPuv fusion protein.

### Package Contents

- 15 µl pET6xHN-N Vector (In-Fusion Ready) [100 ng/µl]
- 15 µl pET6xHN-C Vector (In-Fusion Ready) [100 ng/µl]
- 10 µl pET6xHN-GFPuv Vector [500 ng/µl]
- 15 µl 1.1 kb LacZ-RK Control Insert [25 ng/µl]
- 20 µl 5X In-Fusion HD Enzyme Premix

### Storage Conditions

- Store at -20°C.
- Spin briefly to recover contents.
- Avoid repeated freeze/thaw cycles.

### Expiration Date

- Specified on product label.

### Shipping Conditions

- Dry ice

### Product Documents

Documents for our products are available for download at [takarabio.com/manuals](http://takarabio.com/manuals)

The following documents apply to this product:

- pET Express & Purify Kits User Manual

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#### Takara Bio USA, Inc.

2560 Orchard Parkway, San Jose, CA 95131, USA

U.S. Technical Support: [technical\\_support@takarabio.com](mailto:technical_support@takarabio.com)

United States/Canada  
800.662.2566  
(042623)

Asia Pacific  
+1.650.919.7300

Europe  
+33.(0)1.3904.6880

Japan  
+81.(0)77.565.6999

## pET6xHN-N, pET6xHN-C, and pET6xHN-GFPuv Vector Information

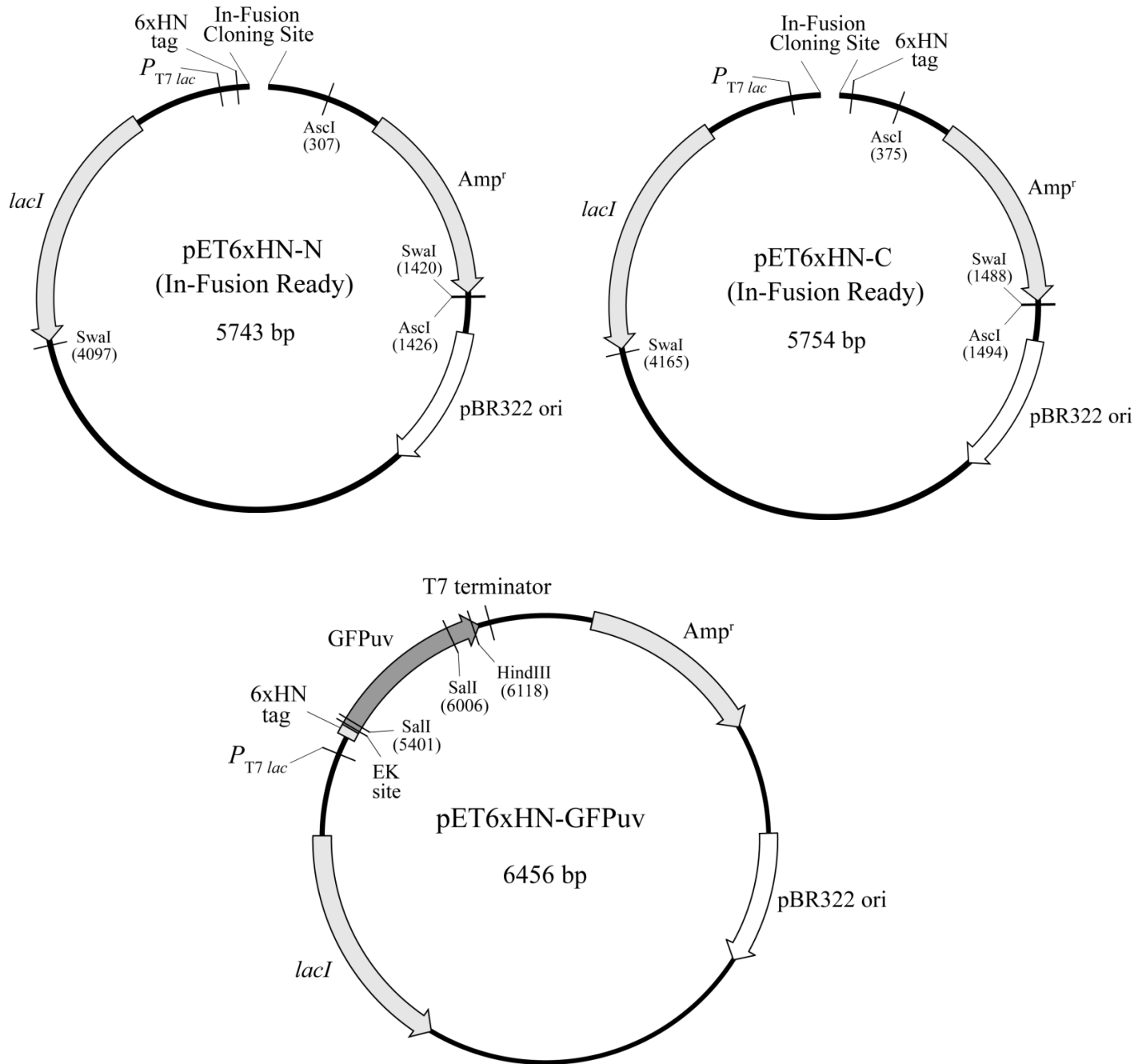


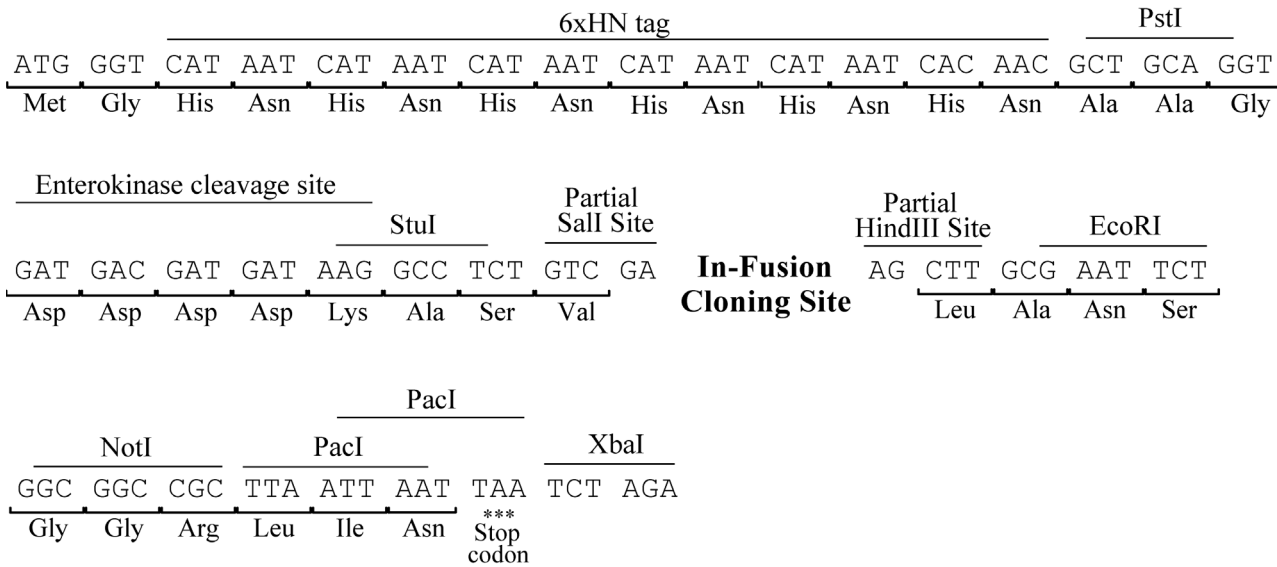
Figure 1. pET6xHN-N (In-Fusion Ready), pET6xHN-C (In-Fusion Ready), and pET6xHN-GFPuv Vector Maps.

# Certificate of Analysis

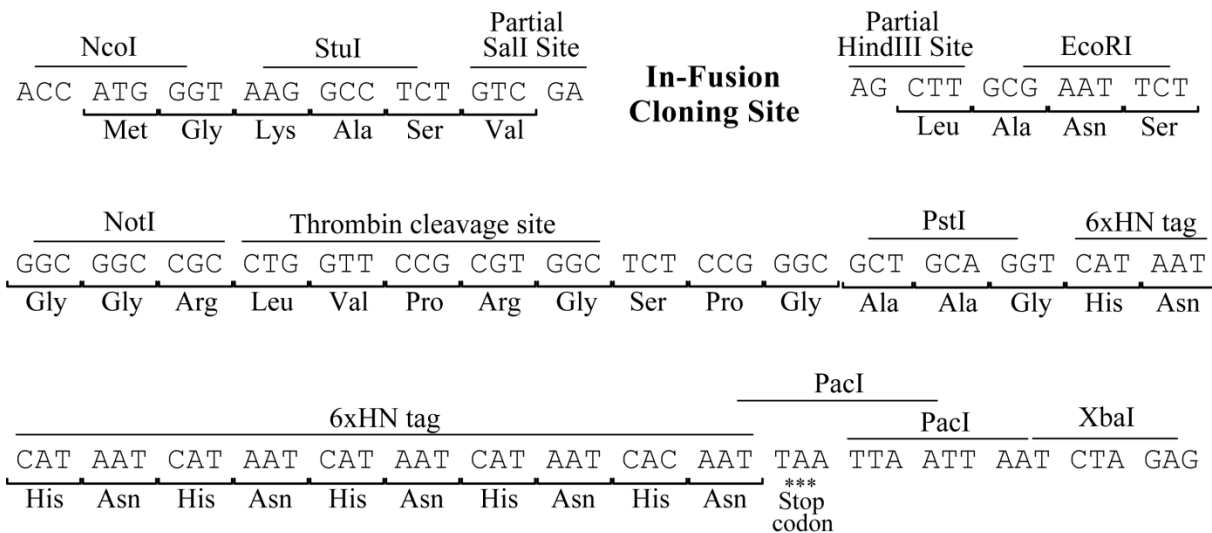
Cat. No. 631433

pET6xHN Expression Vector Set (In-Fusion Ready) [Not sold separately]

Sold as a part of 631428 & 631429



**Figure 2. pET6xHN-N (In-Fusion Ready) multiple cloning site (MCS).** The pET6xHN-N (In-Fusion Ready) vector has been pre-linearized with Sall and HindIII, allowing you to use In-Fusion technology to add an N-terminal 6xHN tag to your protein of interest.



**Figure 3. pET6xHN-C (In-Fusion Ready) multiple cloning site (MCS).** The pET6xHN-C (In-Fusion Ready) vector has been pre-linearized with Sall and HindIII, allowing you to use In-Fusion technology to add a C-terminal 6xHN tag to your protein of interest.

## Description

The vector set contains tightly regulated, highly inducible, bacterial expression vectors (pET6xHN-N and pET6xHN-C) that allow you to express your protein of interest with an N- or C-terminal his tag in *E. coli*. The vectors have been prelinearized with Sall and HindIII for easy In-Fusion cloning of PCR products in-frame with the N- or C-terminal his tag and enterokinase or thrombin cleavage sites. The vectors are based on the pET system vectors developed by William Studier and colleagues at Brookhaven National Laboratories (Dubendorf and Studier 1991; Rosenberg and Studier 1987; Studier and Moffatt 1986; Studier et al. 1990). Derived from pET11 (Dubendorf and Studier 1991), the vectors contain a T7 *lac* hybrid promoter ( $P_{T7 lac}$ ), which combines the strong T7 promoter with the *lac* operator. Basal expression of the protein of interest is repressed by the Lac repressor (*lacI*), which binds to the *lac* operator, preventing expression from the promoter in the absence of IPTG. High-level, IPTG-inducible expression of the protein of interest is driven by the T7 promoter in the presence of T7 RNA polymerase.

Each vector encodes a 6xHN tag composed of 6 repeating His-Asn subunits, (His-Asn)<sub>6</sub>, and either an enterokinase or thrombin cleavage site for subsequent his tag removal. The vectors also contain an ampicillin resistance gene (Amp<sup>r</sup>) and a pBR322 origin of replication, which maintains each vector at a low copy number to further reduce basal levels of the protein of interest.

pET6xHN-GFPuv is a non-linearized control vector that encodes a GFPuv fusion protein containing an N-terminal 6xHN tag and an enterokinase cleavage site. GFPuv is a green fluorescent protein variant optimized for maximal fluorescence when excited by UV light. The vector allows the use of GFPuv fluorescence (excitation and emission maxima at 395 nm and 509 nm, respectively) to monitor protein expression and purification.

## Location of Features

### pET6xHN-N Vector

- T7 terminator: 80–126
- Amp<sup>r</sup> (ampicillin resistance gene; beta-lactamase): 549–1406
- pBR322 origin of replication: 1580–2194
- *lacI* (Lac repressor): 4114–5193 (complementary)
- $P_{T7 lac}$  (T7 *lac* hybrid promoter):
  - T7 promoter: 5580–5596
  - *lac* operator: 5599–5623 (complementary)
- RBS (ribosomal binding site): 5653–5659
- 6xHN tag ([His-Asn]<sub>6</sub>): 5673–5708
- Enterokinase cleavage site: 5718–5732

### pET6xHN-C Vector

- Thrombin cleavage site: 24–41
- 6xHN tag ([His-Asn]<sub>6</sub>): 57–92
- T7 terminator: 148–194
- Amp<sup>r</sup> (ampicillin resistance gene; beta-lactamase): 617–1474
- pBR322 origin of replication: 1648–2262
- *lacI* (Lac repressor): 4182–5261 (complementary)
- $P_{T7 lac}$  (T7 *lac* hybrid promoter):
  - T7 promoter: 5648–5664
  - *lac* operator: 5667–5691 (complementary)
- RBS (ribosomal binding site): 5721–5727

## pET6xHN-GFPuv Vector

- Amp<sup>r</sup> (ampicillin resistance gene; beta-lactamase): 210–1067
- pBR322 origin of replication: 1242–1855
- *lacI* (Lac repressor): 3775–4854 (complementary)
- *P*<sub>T7 *lac*</sub> (T7 *lac* hybrid promoter):
  - T7 promoter: 5241–5257
  - *lac* operator: 5260–5284 (complementary)
- RBS (ribosomal binding site): 5314–5320
- 6xHN tag ([His-Asn]<sub>6</sub>): 5334–5369
- Enterokinase cleavage site: 5379–5393
- GFPuv: 5406–6116
- T7 terminator: 6197–6243

## Additional Information

The In-Fusion cloning sites in pET6xHN-N and pET6xHN-C were created by digesting each vector with Sall and HindIII. As a result, the ends of each vector contain partial Sall and HindIII sites. To maintain the correct reading frame, the nucleotides missing from these partial Sall and HindIII sites should be incorporated into your In-Fusion primers. If you would rather not recreate these restriction sites, be certain not to incorporate the missing nucleotides in your In-Fusion primers.

To express your gene of interest without a 6xHN tag and enterokinase cleavage site, do not use your In-Fusion primers to restore the reading frame. If you want to include a tag of your choice, simply incorporate the tag's sequence into your PCR primers. If you wish to clone your gene of interest into other restriction sites in the MCS, we recommend using the circular versions of these vectors (Takara Bio, Cat. Nos. 631430 & 631431).

Exceptionally pure his-tagged proteins can be obtained with our TALON Co resins (Takara Bio, Cat. Nos. 635501–635504, 635506, 635507, 635509 & 635510) and columns (Takara Bio, Cat. Nos. 635601–635603 & 635606). For routine use, we have a variety of high-capacity His60 Ni resins available (Takara Bio, Cat. Nos. 635659–635664).

## Propagation in *E. coli*

- Suitable host strains for manipulation and propagation: Stellar™ Competent Cells
- Suitable host strains for protein expression: BL21 (DE3) and other DE3 lysogens.
- Selectable marker: plasmid confers resistance to ampicillin (100 µg/ml) in *E. coli* hosts.
- *E. coli* replication origin: pBR322
- Copy number: low

## Excitation and Emission of GFPuv

- Excitation: 395 nm
- Emission: 509 nm

## References

- Dubendorf, J. W. & Studier, F. W. Controlling basal expression in an inducible T7 expression system by blocking the target T7 promoter with lac repressor. *J. Mol. Biol.* **219**, 45–59 (1991).
- Rosenberg, A. H. & Studier, F. W. T7 RNA polymerase can direct expression of influenza virus cap-binding protein (PB2) in *Escherichia coli*. *Gene* **59**, 191–200 (1987).
- Studier, F. W. & Moffatt, B. A. Use of bacteriophage T7 RNA polymerase to direct selective high-level expression of cloned genes. *J. Mol. Biol.* **189**, 113–130 (1986).
- Studier, F. W., Rosenberg, A. H., Dunn, J. J. & Dubendorff, J. W. Use of T7 RNA polymerase to direct expression of cloned genes. *Methods Enzymol.* **185**, 60–89 (1990).

## Quality Control Data

### Plasmid Identity & Purity

- Digestion with the indicated restriction enzymes produced fragments of the indicated sizes on a 0.8% agarose/EtBr gel:

Vector	Enzymes	Fragments
pET6xHN-GFPuv	HindIII	6.5 kb
	SalI	0.6 & 5.9 kb

- Vector identity was confirmed by sequencing.
- A<sub>260</sub>/A<sub>280</sub>: 1.8–2.0

### Functional Assay

In-Fusion HD Enzyme Premix was used to clone 50 ng of the 1.1 kb LacZ-RK Control Insert into both the pET6xHN-N (In-Fusion Ready) vector and the pET6xHN-C (In-Fusion Ready) vector (150 ng each) as described in the User Manual, incubating for 15 min at 50°C. A 2.5 µl aliquot of each mixture was then used to transform Stellar Competent Cells (5 x 10<sup>8</sup> cfu/µg). After 1 hr of growth in 450 µl of SOC Medium, 50 µl of a 1:10 dilution of each transformation mix was plated onto an LB Miller/Amp 100/X-gal IPTG plate. Plates were scored for blue and white colonies after 16–19 hrs.

Negative control cloning reactions lacking the 1.1 kb Control Insert were performed using 150 ng of each vector as described in the User Manual, incubating for 15 min at 50°C. A 2.5 µl aliquot of each negative control reaction was used to transform Stellar Competent Cells (5 x 10<sup>8</sup> cfu/µg). After 1 hr of growth in 450 µl of SOC Medium, each transformation mix was plated onto an LB Miller/Amp 100/X-gal IPTG plate.

It is certified that this product meets the above specifications, as reviewed and approved by the Quality Department.

## pET6xHN Expression Vector Set (In-Fusion® Ready)

### CATALOG NO.

631433

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#### Takara Bio USA, Inc.

2560 Orchard Parkway, San Jose, CA 95131, USA

U.S. Technical Support: [technical\\_support@takarabio.com](mailto:technical_support@takarabio.com)

#### United States/Canada

800.662.2566

#### Asia Pacific

+1.650.919.7300

#### Europe

+33.(0)1.3904.6880

#### Japan

+81.(0)77.565.6999

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