Premix Ex Taq[™] DNA Polymerase Hot Start (Cat.# RR030A)

Application: Amplification of GC-rich DNA Fragments using *TaKaRa LA Taq®* or *Ex Taq®* DNA Polymerase

When amplifying template DNA that is GC-rich, conventional *Taq* DNA Polymerase often does not perform well. The superior amplification efficiency of *TaKaRa LA Taq*[®] and *TaKaRa Ex Taq*[®] allows amplification of difficult DNA targets, such as those with high GC content.

This application example illustrates the ability of these enzymes to amplify a 3 kb fragment that has 70% GC content.

Methods:	VIII 5 mUC110 (s. 2) (h CC with fur amount incorted into the Using III site of mUC110)
Template:	VIII-5-pUC118 (a ~3 kb GC-rich fragment inserted into the <i>Hin</i> d III site of pUC118)
Amplification size:	~3 kb
Primers:	M3-30 (an extended M3 Primer) 5'-CAGTCACGACGTTGTAAAACGACGGCCAGT-3' RV-32 (an extended RV Primer) 5'-GATAACAATTTCACACAGGAAACAGCTATGAC-3' (10 pmol of each primer was used per 50 μl PCR reaction)
dNTP mixture:	7-deaza-dGTP was used instead of dGTP
PCR conditions:	
	94°C, 1 min. ↓ 98°C, 20 sec. 68°C, 5 min 30 cycles
Thermal cycler:	TaKaRa PCR Thermal Cycler 480*
Enzymes:	<i>TaKaRa LA Taq® –</i> LA PCR Kit Ver. 2.1 (Cat.# RR013A) <i>TaKaRa Ex Taq® – Premix Ex Taq</i> ™ DNA Polymerase Hot Start Version (Cat.# RR030A)

Results:



Figure 1. Amplification of a GC-rich fragment using various DNA polymerases. PCR products were amplified using either conventional *Taq* (lane 1), *Ex Taq*[®] (lane 2), or *LA Taq*[®] (lane 3) and were run on a 1% L03 agarose gel (8 μ l PCR reaction per lane). Lanes M1 and M2 contain molecular weight markers (λ -*Hind* III digest and λ -*Eco*T14 digest, respectively).

*Not available in all geographic locations. Check for availability in your region.

TaKaRa

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Using 7-deaza-dGTP instead of dGTP in the PCR reaction can improve amplification of GC-rich templates. In addition, further optimization using a mixture of dGTP and 7-deaza-dGTP (or dITP) can also improve amplification efficiency (Figure 2).



Figure 2. Amplifications of a GC-rich fragment using various ratios of dGTP and 7-deazadGTP or dITP. PCR products were amplified using either *LA Taq*[®] (lanes 1–3) or *Ex Taq*[®] (lanes 4–6) and were run on a 1% L03 agarose gel (10 μ l). The PCR reactions contained various combinations of dGTP, dITP, and 7-deaza-dGTP as indicated in the table above. Lane M contains a pHY molecular weight marker.

Lane 1: dGTP:dITP = 3:1 Lane 2: dGTP 7-deaza-dGTP = 3:1 Lane 3: 7-deaza-dGTP alone

Lane 4: dGTP:dITP = 3:1 Lane 5: dGTP:7-deaza-dGTP = 3:1 Lane 6: 7-deaza-dGTP alone

Conclusions:

A GC-rich fragment that was not efficiently amplified using conventional *Taq* could be successfully amplified with both *TaKaRa Ex Taq*[®] and *TaKaRa LA Taq*[®] DNA polymerases.

Amplification of GC-rich target sequences can be improved by replacing dGTP with dITP or 7-deaza-dGTP. Depending on fragment to be amplified, the optimal ratio of dGTP to 7-deaza-dGTP (or dITP) may vary. Therefore, we recommend evaluating various ratios to determine the ideal conditions.